

1 **LamPORE SARS-CoV-2 diagnosis and genotyping: a preliminary report.**

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13 **Dear Editor.**

14 Direct diagnosis of COVID-19 is mainly based on the reverse-transcription PCR (RT-  
15 PCR) detection of the SARS-COV-2 RNA (1) (2). The LamPORE process has been  
16 recently developed for detecting the viral genome by the reverse-transcription loop-  
17 mediated amplification (RT-LAMP) and sequencing the RT-LAMP product on  
18 LamPORE instrument (Oxford Nanopore, Oxford, UK) (3). Here, 264 nasopharyngeal  
19 swabs routinely submitted to BGI's Real-Time Fluorescent RT-PCR kit (BGI, Wuhan,  
20 China), (61 positive and 203 negative samples) at the IHU Méditerranée Infection,  
21 Marseille, France, were investigated in parallel by the LamPORE procedure. Briefly, the  
22 LamPORE procedure combines an isothermal amplification step for 30 minutes at 65°C,  
23 performed in a 50- $\mu$ L volume containing 20  $\mu$ L RNA on Kingfisher instrument, using  
24 MagMax™ Viral/Pathogen Kit (ThermoFisher, St. Austin, USA), 5  $\mu$ L priming Mix and  
25 25  $\mu$ L of LamPORE master Mix, targeting three SARS-COV-2 genes: ORF1a, the  
26 envelope (E) and nucleocapsid (N) genes in addition to the human actin mRNA as  
27 internal control. Further, 2  $\mu$ L of each amplified sample were engaged in LamPORE  
28 library preparation as previously described (3) and the library was sequenced on  
29 LamPORE instrument for one hour. LamPORE assays incorporated water as negative  
30 control, provided by the manufacturer. LamPORE diagnosis results interpretation  
31 positive and negative tests automatically generated in a pdf file as follows: positive test  
32 when the sum of reads generated by the three targets  $\geq 50$ ; negative test when number  
33 of reads  $< 20$ ; inconclusive test when number of reads between 20 and 50. Strictly using  
34 the interpretation algorithm proposed by the manufacturer, all the 61 nasopharyngeal  
35 swabs routinely positive for SARS-COV-2 were found positive by LamPORE (100%

36 sensitivity) but 23/203 of RT-PCR negative nasopharyngeal swabs detected positive by  
37 LamPORE (88.7% specificity). Analyzing these 23 discordant results, we observed that  
38 for a result to be consistent with RT-PCR, the sum of the 3 targets reads > 50 and the  
39 multiplication of the N (reads) of the 3 targets > 0 was required. Applying this new  
40 interpretation rule allowed to achieve 100% sensitivity and 100% specificity, in the 61-  
41 sample collection we investigated. This new interpretation rule consisted in eliminating  
42 false positives by applying the “=SI(ET((C2+D2+E2>50);(C2\*D2\*E2>0));"pos";"neg")”  
43 formula directly in the Excel file. In a second step, we used sequences to detect SARS-  
44 COV-2 genotype 4, the most prevalent genotype in the Marseille area in the considered  
45 period. Indeed, LamPORE theoretically generates substitutions G28,975T and  
46 G29,399A in the nucleocapsid gene, among the 13 mutations specific for genotype 4  
47 (Figure 1). Here, LamPORE detected two genotype 4, based on two sequences  
48 exhibiting only the G28,975T mutation and one sequence exhibiting the mutation  
49 G29,399A.

50         Although preliminary, data here reported confirm that LampORE is an  
51 appropriate method for the rapid direct detection of SARS-CoV-2 RNA in  
52 nasopharyngeal swabs, with a capacity of 480 tests per hour, depending on adoption of  
53 the interpretation rule here reported; and offer promise for one-shot genotyping of  
54 SARS-CoV-2 depending on further experimental improvements, offering a new  
55 alternative way for SARS-COV-2 detection and genomic surveillance.

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63 **Ethics**

64 Only nasopharyngeal residual fluid left from standard-of-care clinical laboratory testing  
65 was used. All specimens had been referred to our laboratory for diagnostic purposes  
66 between September 1<sup>st</sup> and November 1<sup>st</sup>, 2020. The study was validated by our  
67 Institute's Ethical Committee under number 2020-016603.

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75 **Conflicts of interest**

76 Reagents for the LamPORE instrument have been provided by Oxford Nanopore  
77 Technology, Oxford, UK. Nevertheless, the supplier did not interfere in experimental  
78 plan, data interpretation, manuscript preparation and submission.

79 **Author's contribution.**

80 MM, contributed to experimental design, realization of the work, data analysis,  
81 interpretation, and writing. AH, collected samples, data analysis, and writing. BL,  
82 organization of the work. DJ, Bioinformatics data analysis. BM, experimental design.  
83 PEF and DM, contributed to critically reviewing the manuscript, data interpretation,

84 coordinated and directed the work. All authors declare that they have read and  
85 approved the manuscript.

86 **References**

- 87 1. Chakraborty C, Sharma AR, Sharma G, Bhattacharya M, Lee SS. SARS-CoV-2  
88 causing pneumonia-associated respiratory disorder (COVID-19): Diagnostic and  
89 proposed therapeutic options. *Eur Rev Med Pharmacol Sci* [Internet]. 2020;  
90 24(7):4016–26. Available from: <https://pubmed.ncbi.nlm.nih.gov/32329877/>
- 91 2. Zheng Z, Yao Z, Wu K, Zheng J. The diagnosis of SARS-CoV2 pneumonia: A  
92 review of laboratory and radiological testing results [Internet]. Vol. 92, *Journal of*  
93 *Medical Virology*. John Wiley and Sons Inc; 2020. p. 2420–8. Available from:  
94 <https://pubmed.ncbi.nlm.nih.gov/32462770/>
- 95 3. James P, Stoddart D, Harrington ED, Beaulaurier J, Ly L, Reid SW, et al.  
96 LamPORE: rapid, accurate and highly scalable molecular screening for SARS-  
97 CoV-2 infection, based on nanopore sequencing. *medRxiv*;  
98 2020.08.07.20161737. Available from:  
99 <https://doi.org/10.1101/2020.08.07.20161737>

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103 **Figure 1.** LamPORE diagnosis strategy for SARS-COV-2 detection and genotyping.  
104 LAMP amplification and library preparation were followed by library sequencing on the  
105 LamPORE instrument. A genotyping step was added to specifically detect SARS-COV-  
106 2 genotype 4.

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